

```

seed = -1
treefile = MyTree.tre
Imapfile = MyImap.txt
species&tree = 3 A B C
          2 1 0
((A #0.05, B) :0.005 #0.001, C #0.05) :0.01 #0.01;
loci&length = 1000000 50
migration = 1
      C A 1.0

```

**Fig. S1:** BPP control file (MCcoal-ABC-mscm.ctl) for simulating  $10^6$  gene trees under the MSC-M model of Fig. 10. The species tree is specified using the Newick notation, with numbers after colon ‘:’ indicating node ages ( $\tau$ ) while those after ‘#’ are population sizes ( $\theta$ ). Note that species *C* is needed in the specification of the tree and migration rate, but no sequences are sampled from *C* when gene trees are simulated. The sequence length is unused since no sequence data are simulated. Run BPP with the command

```
bpp --simulate MCcoal-ABC-mscm.ctl
```

```

Number of species in starting delimitation: 5
(X, ((A, B), (C, D)));

*** Iteration 1 ***

Estimated tau and theta parameters:
      theta           tau
X     0.0100 (0.0097, 0.0103)
A     0.0111 (0.0095, 0.0126)
B     0.0242 (0.0193, 0.0294)
C     0.0262 (0.0209, 0.0318)
D     0.0106 (0.0093, 0.0120)
XABCD 0.0330 (0.0321, 0.0340) 0.0106 (0.0104, 0.0108)
ABCD 0.0442 (0.0434, 0.0450) 0.0005 (0.0005, 0.0006)
AB    0.0068 (0.0041, 0.0095) 0.0004 (0.0003, 0.0004)
CD    0.0059 (0.0037, 0.0084) 0.0004 (0.0004, 0.0005)

Proposal results:
Node pair      gdi 1   gdi 2   merge accepted?
'A', 'B'       0.068   0.032   True
'C', 'D'       0.031   0.075   True

Number of species after iteration 1: 3
(X, (AB, CD));

*** Iteration 2 ***

Estimated tau and theta parameters:
      theta           tau
X     0.0100 (0.0097, 0.0103)
AB    0.0203 (0.0190, 0.0216)
CD    0.0204 (0.0191, 0.0217)
XABCD 0.0331 (0.0322, 0.0340) 0.0106 (0.0104, 0.0108)
ABCD 0.0433 (0.0425, 0.0441) 0.0006 (0.0005, 0.0006)

Proposal results:
Node pair      gdi 1   gdi 2   merge accepted?
'AB', 'CD'     0.054   0.054   True

Number of species after iteration 2: 2
(X, ABCD);

*** Iteration 3 ***

Estimated tau and theta parameters:
      theta           tau
X     0.0100 (0.0097, 0.0103)
ABCD 0.0434 (0.0427, 0.0440)
XABCD 0.0331 (0.0321, 0.0340) 0.0106 (0.0104, 0.0108)

Proposal results:
Node pair      gdi 1   gdi 2   merge accepted?
'X', 'ABCD'    0.879   0.388   False

Number of species after iteration 3: 2
(X, ABCD);

Final delimitation reached.

```

**Fig. S2:** Screen output from running the hierarchical merge algorithm to analyze the simulated dataset of Fig. 4.

```

# Note species are renamed as follows:
# Po = polyzona
# Ab = abnorma
# Mi = micropholis
# An = annulata
# Ge = gentilis
# Tr = triangulum
# El = elapsoides

# Note that species are renamed as follows:
# gir_ang = giraffa+angolensis
# tip_tho = tippelskirchi+thornicrofti
# cam_rot_ant = camelopardalis+rothschildi+antiquorum
# per = peralta
# ret = reticulata

# output
output_directory = res_giraffe_merge

# input files
Imapfile = Imap_Giraffe.txt
seqfile = MSA_Giraffe.txt

# guide tree
guide_tree = ((gir_ang, tip_tho), ((cam_rot_ant, per), ret));

# migration events and priors
migration = {
    ret <-> tip_tho,
    ret <-> cam_rot_ant,
}
migprior = 0.1 10

# hierarchical algorithm settings
mode = merge
gdi_threshold = <=0.3, <=1.0

# BPP MCMC settings
threads = 8
burnin = 50000
nsample = 200000

# Note species are renamed as follows:
# Po = polyzona
# Ab = abnorma
# Mi = micropholis
# An = annulata
# Ge = gentilis
# Tr = triangulum
# El = elapsoides

# output
output_directory = res_milksnake_merge

# input files
Imapfile = Imap_Lampropeltis.txt
seqfile = MSA_Lampropeltis.txt

# guide tree
guide_tree = (((Mi, (Po, Ab)), (An, (Ge, Tr))), El);

# migration events and priors
migration = {
    Po <-> Ab,
    Po <-> An,
    An <-> Ge,
    Ge <-> Tr,
    Ge <-> El,
    Tr <-> El,
}
migprior = 0.1 10

# hierarchical algorithm settings
mode = merge
gdi_threshold = <=0.3, <=1.0

# BPP MCMC settings
threads = 8
burnin = 50000
nsample = 200000

```

**Fig. S3:** HHSD control file for the merge algorithm in analysis of the giraffe data. The control file for the split algorithm is the same except for the hierarchical-algorithm settings: mode = split and gdi\_threshold = >0.7. The results are in Fig. 13c&d.

**Fig. S4:** HHSD control file for the merge analysis of the milksnakes data. The control file for the split algorithm is the same except for the hierarchical-algorithm settings: mode = split and gdi\_threshold = >0.7. The results are in Fig. 14c&d.

```

# output
output_directory = # will be set from the command line

# input files
Imapfile = # will be set from the command line
seqfile = trigentalt.txt

# guide tree
guide_tree = ((Ge, Tr), Al);

# migration events and priors
migration = { Ge <-> Tr }
migprior = 0.1 10

# hierarchical algorithm settings
mode = merge
gdi_threshold = <0.3

# BPP MCMC settings
threads = 8
burnin = 200000
nsample = 500000

```

**Fig. S5:** HHSD control file for the analysis of the milk-snake data under the five delimitation hypotheses with arbitrary East-West divide of *gentilis* and *triangulum* samples (Fig. 14e). The Imapfile and output\_directory parameters are left empty, as they will be provided at the command line (Fig. S6), to ensure that they correspond to the delimitation hypotheses being tested. *L. alterna* (Al) is used as an outgroup, following Chambers and Hillis (2020).

```

hhsd --cfile cf_milksnake_EW.txt --cfpor \
Imapfile = 1alt.Imap.txt, output_directory = res_EW_1

hhsd --cfile cf_milksnake_EW.txt --cfpor \
Imapfile = 2alt.Imap.txt, output_directory = res_EW_2

hhsd --cfile cf_milksnake_EW.txt --cfpor \
Imapfile = 3alt.Imap.txt, output_directory = res_EW_3

hhsd --cfile cf_milksnake_EW.txt --cfpor \
Imapfile = 4alt.Imap.txt, output_directory = res_EW_4

hhsd --cfile cf_milksnake_EW.txt --cfpor \
Imapfile = 5alt.Imap.txt, output_directory = res_EW_5

```

**Fig. S6:** Shell script used to iterate through the five East-West delimitation hypotheses for the milksnakes (Fig. 14e). The -cfpor (control file parameter override) flag is used to override parameters of the control file via the command-line interface, setting the Imap file to match the delimitation hypothesis, and specifying the output directory for each analysis.

```

# Note species are renamed:
# PEL = pelastes
# OZK = ozark
# MEG = megalotis
# LIT = ouachita
# SOL = solis
# AQU = aquilensis

# output
output_directory = res_sunfish_merge

# input files
Imapfile = Imap_Sunfish.txt
seqfile = MSA_Sunfish.txt
phase = 1 # alignment is phased

# guide tree
guide_tree = (((((PEL, OZK), MEG), LIT), SOL), AQU);

# migration
migration = {
    MEG -> PEL,
    MEG -> SOL,
    MEG -> OZK
}
migprior = 0.1 10

# hierarchical algorithm settings
mode = merge
gdi_threshold = <=0.3, <=1.0

# BPP MCMC settings
threads = 12
burnin = 500000
nsample = 200000

```

**Fig. S7:** HHSD control file for the merge analysis of the sunfish data (Fig. 15). The control file for the split algorithm is the same except for the hierarchical-algorithm settings: mode = split and gdi\_threshold = >0.7.

**Table S1:** Rate matrix for Markov chain describing transitions between states in multispecies coalescent with migration model with two populations ( $A$  and  $B$ ) and three sequences ( $a_1, a_2$ , and  $b$ ).

	$AA$	$AAB$	$ABA$	$ABB$	$BAA$	$BAB$	$BBA$	$BBB$	$A_{a_1}A$	$A_{a_2}A$	$A_bA$	$B_{a_1}B$	$B_{a_2}B$	$B_bB$	$A_{a_1}B$	$A_{a_2}B$	$A_bB$	$AB_{a_1}$	$AB_{a_2}$	$AB_b$	$A B$		
$AA$	.	$\varpi_{BA}$	$\varpi_{BA}$	.	$\varpi_{BA}$	$\varpi_{BA}$	$\varpi_{BA}$	$\varpi_{BA}$	$c_A$	$c_A$	$c_A$	.	.	.	.	.	.	.	$c_A$	$c_A$	$c_A$	.	
$AAB$	$\varpi_{AB}$	.	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	.	.	.	.	.	.	.	.	.	$c_A$	$c_A$	$c_A$	.	
$ABA$	$\varpi_{AB}$	.	.	.	$\varpi_{BA}$	.	.	$\varpi_{BA}$	.	.	.	.	.	.	.	.	.	.	$c_A$	$c_A$	$c_A$	.	
$ABB$	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	.	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$c_B$	.	.	$c_B$	$c_B$	$c_B$	.	
$BAA$	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	$c_B$	.	.	$c_B$	$c_B$	$c_B$	.
$BAB$	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$c_B$	.	.	$c_B$	$c_B$	$c_B$	.											
$BBA$	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$c_B$	.	.	$c_B$	$c_B$	$c_B$	.											
$BBB$	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$c_B$	.	.	$c_B$	$c_B$	$c_B$	.											
$A_{a_1}A$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{BA}$	.	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	
$A_{a_2}A$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{BA}$	.	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	
$A_bA$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{BA}$	.	.	$\varpi_{BA}$	.	$\varpi_{BA}$	.	
$B_{a_1}B$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$B_{a_2}B$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$B_bB$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$A_{a_1}B$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$A_{a_2}B$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$A_bB$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$AB_{a_1}$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$AB_{a_2}$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$AB_b$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	$\varpi_{AB}$	.	.	$\varpi_{AB}$	.	$\varpi_{AB}$	.	
$A B$	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	

Note.—  $\varpi_{AB} = 4M_{AB}/\theta_B = m_{AB}/\mu$  and  $\varpi_{BA} = 4M_{BA}/\theta_A = m_{BA}/\mu$  are mutation-scaled migration rates, and  $c_A = 2/\theta_A$  and  $c_B = 2/\theta_B$  are the coalescent rates, when one time unit is the expected time to accumulate one mutation per site. The state of the chain is given by the population IDs ( $A$  or  $B$ ) and sequence IDs ( $a_1, a_2, b$ ). For example the initial state  $A_{a_1}A_{a_2}B_b$  means that the three sequences  $a_1, a_2$ , and  $b$  are in populations  $A$ ,  $A$ , and  $B$ , respectively. States with three sequences assumed to be in the order  $a_1, a_2, b$ , so that the sequence IDs are suppressed. Thus  $A_{a_1}A_{a_2}B_b$  is ‘ $AAB$ ’. State  $A_{a_1}a_2B_b$  is ‘ $AAB$ ’. State  $A_{a_1}a_2B_b$  means that two sequences remain in the sample, with the ancestor of sequences  $a_1$  and  $a_2$  is in population  $A$  while sequence  $b$  is in population  $B$ . This is abbreviated ‘ $AB_b$ ’, with the sequence ID ‘ $a_1a_2$ ’ suppressed. ‘ $A|B$ ’ is an absorbing state in which only one sequence remains in the sample, in either  $A$  or  $B$ , after two coalescent events have occurred. From [Leaché et al. \(2019\)](#).

**Table S3:** Hierarchical merge algorithm under the MSC-M model allowing the merge of non-sister lineages applied to data simulated under the model of Fig. 4a

Pair	$gdi_{ij}$			$gdi_K$		
	$aab$	$abb$	Decision	$aab$	$abb$	Decision
<b>Using cutoff: <math>gdi &lt; 0.2</math></b>						
Iteration 1: merge of nonsister lineages $A$ and $B$ is accepted						
$X-A$	0.876	0.433	rejected	0.861	0.428	rejected
$A-B$	0.078	0.028	<b>accepted</b>	0.322	0.284	rejected
$A-C$	0.126	0.075	rejected	0.399	0.365	rejected
$B-C$	0.052	0.052	rejected	0.351	0.350	rejected
$A-D$	0.153	0.159	rejected	0.434	0.439	rejected
$B-D$	0.082	0.138	rejected	0.387	0.424	rejected
$C-D$	0.031	0.090	rejected	0.354	0.392	rejected
Iteration 2: merge of nonsister lineages $C$ and $D$ is accepted						
$X-AB$	0.874	0.402	rejected			
$AB-C$	0.060	0.066	rejected			
$AB-D$	0.088	0.148	rejected			
$C-D$	0.031	0.088	<b>accepted</b>			
Iteration 3: merge of nonsister lineages $AB$ and $CD$ is accepted						
$X-AB$	0.877	0.388	rejected			
$AB-CD$	0.073	0.070	<b>accepted</b>			
Iteration 4: merge of sister lineages $X$ and $ABCD$ is rejected						
$X-ABCD$	0.879	0.388	rejected			
Final model: two species: $X, ABCD$ five species: $X, A, B, C, D$						
<b>Using cutoff: <math>gdi &lt; 0.7</math></b>						
Iteration 1: merge of sister lineages $X$ and $A$ is accepted						
$X-A$	0.876	0.433	<b>accepted</b>	0.861	0.428	<b>accepted</b>
$A-B$	0.078	0.028	rejected	0.322	0.284	rejected
$A-C$	0.126	0.075	rejected	0.399	0.365	rejected
$B-C$	0.052	0.052	rejected	0.351	0.350	rejected
$A-D$	0.153	0.159	rejected	0.434	0.439	rejected
$B-D$	0.082	0.138	rejected	0.387	0.424	rejected
$C-D$	0.031	0.090	rejected	0.354	0.392	rejected
Iteration 2: merge of sister lineages $XA$ and $B$ is accepted						
$XA-B$	0.132	0.237	<b>accepted</b>	0.357	0.433	<b>accepted</b>
$XA-C$	0.150	0.214	rejected	0.405	0.450	rejected
$B-C$	0.085	0.039	rejected	0.375	0.344	rejected
$XA-D$	0.159	0.270	rejected	0.438	0.512	rejected
$B-D$	0.121	0.127	rejected	0.413	0.417	rejected
$C-D$	0.030	0.081	rejected	0.353	0.386	rejected
Iteration 3: merge of sister lineages $XAB$ and $C$ is accepted						
$XAB-C$	0.048	0.142	<b>accepted</b>	0.292	0.362	<b>accepted</b>
$XAB-D$	0.064	0.214	rejected	0.374	0.475	rejected
$C-D$	0.036	0.093	rejected	0.356	0.394	rejected
Iteration 4: merge of sister lineages $XABC$ and $D$ is accepted						
$XABC-D$	0.024	0.169	<b>accepted</b>	0.317	0.418	<b>accepted</b>
Final model: one species: $XABCD$ one species: $XABCD$						

**Table S2:** Hierarchical merge algorithm under the MSC model applied to data simulated under the model of Fig. 4a

Pair	$gdi_A$ (aab data)	$gdi_B$ (abb data)	Decision
<b>Using the cutoff: <math>gdi &lt; 0.2</math></b>			
Iteration 1: merging $A-B$ into $AB$ , $C-D$ into $CD$			
$A-B$	0.067	0.033	<b>accepted</b>
$C-D$	0.031	0.075	<b>accepted</b>
Iteration 2: merging $AB-CD$ into $ABCD$			
$AB-CD$	0.054	0.054	<b>accepted</b>
Iteration 3: merging $X-ABCD$ into $XABCD$			
$X-ABCD$	0.879	0.388	rejected
Final model: two species: $X, ABCD$			
<b>Using the cutoff: <math>gdi &lt; 0.7</math></b>			
Iteration 1: same as above			
Iteration 2: same as above			
Iteration 3: merging $X-ABCD$ into $XABCD$			
Final model: one species: $XABCD$			

Note.— The guide tree,  $((A, B), (C, D)), X$ , is inferred under the MSC model with no gene flow (Fig. 4b). Under MSC model with no gene flow,  $gdi_j$  and  $gdi_K$  are equivalent (equation 2). A merge (between sister lineages) is accepted if either  $gdi_A$  or  $gdi_B$  is less than the cutoff, with multiple merges allowed in each iteration.

Note.—  $gdi$  values are calculated under the MSC-M model by simulating gene trees using posterior means of parameters (which are very similar to the posterior means of  $gdi$  as the dataset is large). The MSC-M model of Fig. 4a is used as the guide tree (initial delimitation).